# Isolation and culture of endothelial cells from mouse aorta

## Preparation of aorta rings

- 1. Euthanatize mice by CO<sub>2</sub> asphyxiation followed by cervical dislocation.
- 2. Wipe off skin with 70% isopropanol, and fix mice on an operation table.
- 3. Cut off tissue to open chest and abdomen, and expose aorta.
- 4. Dissect thoracic aorta out, and put it into PBS washing solution in 35-mm petri dishes.
- 5. Under a stereomicroscope, remove connective tissue around adventitia.
- 6. Transfer agrta to new petri dishes, and cut it to  $1\sim2$  mm cross-sectional slices (rings) using an iris scissors.

## Implantation of aorta rings in matrix

- 1. Coat new petri dishes with matrix by adding 50 µl diluted Matrigel solution.
- 2. Allow Matrigel to polymerize at room temperature for 20 min.
- 3. Place a rings on the Matrigel layer, and cover the a rta tissue with a few drops of Matrigel.
- 4. Allow Matrigel to polymerize at room temperature for 20 min.
- 5. Wash once with *pre-warmed* PBS.
- 6. Add *pre-warmed* culture medium, and allow endothelial cells (EC) to sprout and grow for 5~7 days at 37 °C in a humid incubator chamber with 5% CO<sub>2</sub>.

# Isolation of sprouting EC

- 1. When a large number of cells sprouting from a re observed, remove a rings from Matrigel with a sterile fine-tip forceps under a stereomicroscope.
- 2. Wash remaining Matrigel and cells with PBS twice.
- 3. Add dispase solution (0.1 ml/cm<sup>2</sup>), and incubate for 20 min at 37 °C in a humid incubator chamber.
- 4. Pipette solution up and down, and transfer solution into a 15-ml tube.
- 5. Add *pre-chilled* 10 ml culture medium, and centrifuge at 500 X g for 10 min at 4 °C.
- 6. Discard supernate, and suspend the cell pellets with 1 ml culture medium.

### **EC** culture

- 1. Coat a dish/flask with fibronectin (0.2 ml/cm<sup>2</sup>) at 4 °C overnight, and wash twice with PBS.
- 2. Add EC suspension and culture medium to the dish/flask.
- 3. Change medium every 3 days and split cells when reaching ~90% confluence.

#### **Appendix**

**PBS washing buffer** PBS with antibiotics (50 μg/ml penicillin and strptomycin)

**Matrigel solution** Add pre-chilled PBS into Matrigel (BD Biosciences, Cat# 356231) at 1:1 on ice. **Dispase solution** Dispase I (Sigma Cat# D4818, 2 U/ml) in PBS.

Fibronectin solution Fibronectin (Sigma Cat# F1141, 10 µg/ml) in PBS.

**Culture medium** Ham's F12/DMEM medium with 5~10% FBS (5% for long-term culture), 50 μg/ml ECGS (endothelial cell growth supplement, made from bovine hypothalami or Sigma Cat# E2759), 10 U/ml heparin (Sigma Cat# 9399), 50 μg/ml penicillin, 50 μg/ml streptomycin.

